



Molecular physiology of manganese in insects

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Manganese is an essential element for maintaining life. Overexposure to the metal, however, can be toxic to organisms. Given the significant function of manganese in insects, agriculture, and human disease, as well as in the healthy ecology of the planet, the biological activities of manganese in insects needs consideration. Because of the role of manganese as a cofactor for essential enzymes present in different organelles, both over and underexposure to manganese has a multifaceted effect on organisms. At the physiological level, the effects of insect exposure to the metal on enzymatic activities and consequent alteration of insect behaviors are best explained through the metal's role in modulating the dopaminergic system. Despite numerous examples that alterations in manganese homeostasis have profound effects on insects, the cellular mechanisms that ensure homeostasis of this essential metal remain presently unknown, calling for further research in this area.

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Background

This literature review covers the physiological role of manganese in insects, at the cellular and tissue level, as well as the metal's impact on the whole animal, which can result in behavioral changes and affect ecological interactions. Insects have often been used as a model to study the effects of manganese overexposure, but only few studies have determined physiological consequences of manganese deficiency.

Manganese (Mn) can exist in a variety of oxidation states as Mn^{2+} , Mn^{4+} , and Mn^{7+} [1**]. Most experimental studies have examined the effects of Mn as $MnSO_4$ or $MnCl_2$ which are readily dissociated in water to Mn^{2+} . The presence of $MnSO_4$ in agriculture sprays and soil

enrichment has increased the exposure of many species to this metal. Mn is essential for photosynthesis in plants (as part of the metal cluster supporting water oxidation in photosystem II, [2]) and promotes plant growth and health. A protein analysis in *Arabidopsis* revealed that 398 enzymes are predicted to contain Mn in a metal binding site, 20% of which have been verified experimentally to use Mn as a cofactor [3], explaining its use for agricultural crops. Application of $MnSO_4$ affects not only the insect populations directly feeding on plants, but also the soil runoff into water sources, including downstream and river-marine deltas. Relatively high levels of $MnSO_4$ are used to increase agricultural production, and these levels amplify throughout the ecosystem (e.g. blueberries, [4]). The effluents of mining and coal excavation also introduces Mn to the environment, serving as another source for exposure to insects. The environment and food web can amplify Mn concentration indirectly in insects in various ways as following: through exposure of pollen, thus affecting honey produced by bees; water sources, often containing developing mosquito larvae or other water borne insects; the bacteria and fungi in the soil that are fed on by termites; as well as other animal food sources used by carnivorous insects.

Dependence of biochemical reactions on Mn^{2+} ions

As in plants, Mn is essential to insects and other animals since it serves as a cofactor for numerous important cellular enzymes and organelle function (Table 1). This includes three mitochondrial enzymes: superoxide dismutase (SOD2; also referred to as manganese superoxide dismutase or MnSOD), arginase, and glutamine synthase [5**]; it also is a cofactor for glycosyltransferases in the Golgi [5**]. The DNA sequences encoding for MnSODs are conserved between animal species [6], suggesting that Mn serves as a cofactor for these enzymes throughout the animal kingdom [7]. MnSOD function has been studied in several insects, including the desert beetle *Micordera punctipennis* [8*], the Asiatic rice borer *Chilo suppressalis* (Walker) (Lepidoptera: Crambidae [9**], the fruit fly *Drosophila melanogaster* [10*], and the insect cell line Sf-9 [11]. Since it is known that MnSOD is important to reduce oxidative stress, a lack of Mn decreases the ability of insects to withstand bacterial and viral infections [11]. Another important enzyme which requires Mn is the NAD-linked "malic" enzyme in skeletal muscle, studied in the tse-tse fly *Glossina morsitans* [12]. It is also important to note that temperature and pesticides may alter the expression of Mn-associated enzymes. Methyl paraoxonase activity was shown to increase in the tufted apple bud moth *Platyota idaеusalis* after exposure to Mn and in

Table 1

The action of Mn on various enzymes and metabolic processes

Insect	Enzyme/compound	Action of Mn	Citation
Aphids (<i>Myzus persicae</i>)	Catalase	Increased activity	[14]
	Glucose-6-phosphate dehydrogenase	Increased activity	
	Superoxide dismutase	Increased activity	
	Peroxidase malondialdehyde (biomarker of oxidative stress)	Increased activity	
		Decreased activity	
	Digestive enzymes:		
	α-amylase	Inhibited activity	
	Trypsin	Inhibited activity	
	Chymotrypsin	Inhibited activity	
	Elastase	Inhibited activity	
Bumble-bee (<i>Bombus spp.</i>)	Fructose 1,6-diphosphatase	Inhibited activity in body wall muscle	[15]
<i>Drosophila melanogaster</i>	Beta4-galactosyltransferase	Increased activity	[16]
	Manganese-dependent Metallopeptidase	Increased activity	[17]
	GTP cyclohydrolase	Inhibited activity	[18]
	CMP-sialic acid synthetase	Increased activity	[19]
	Sugar donor enzyme for sialylation		
House fly (<i>Musca domestica</i>)	Glycerophosphodiesterase	Increased activity	[20]
German cockroach & tobacco budworm	Methyl paraoxonase	No effects	[13]
Mosquito (<i>Aedes aegypti</i>)	Ribonuclease	Increase activity	[21]
	Farnesyl pyrophosphate Synthetases I and II	Increased activity	[22]
Silkworm (<i>Bombyx mori</i>)	Catalase for cinnabarinic acid	Increased activity	[23]
	Cyclic GMP phosphodiesterase	Increased activity	[24]
	Adenylate cyclase	Increased activity	[25]
	Pupal fat body		
Termite (<i>Neotermes koshunensis</i>)	β-glucosidase	Increased activity	[26]
Tufted apple bud moth (<i>Platynota idaeusalis</i>)	Methyl paraoxonase	Increased activity	[13]

building resistance to the pesticide azinphosmethyl [13]. Using current approaches to analyze total mRNA expression (RNA-Seq) for whole animals of small size or even single cells within defined tissues with single cell RNA-Seq, one could assess the effects on gene expression with depletion or over exposure of Mn in order to have a more complete understanding in the protein connectome by such transcriptomic evidence.

Effects of Mn at the cellular level

Mn must be transported into the cell. The *Makvolio* (*Mvl*) gene encodes a proton-coupled metal ion transporter in the SLC11 family which transports Fe^{2+} and Mn^{2+} into cells [27]. Mutations in this gene in *Drosophila melanogaster* affects taste sensation [28*]. Taste in *Drosophila* is dependent on the dopaminergic system within the CNS [29*], so it is of interest to know how Mn may affect the dopaminergic neurons. Additionally, altered neuronal *Mvl* expression in bees affects social behavior [30**]. It is worth mentioning that a high-affinity Mn transport

system has been hypothesized in *Drosophila* [5**], although the specific transporter involved has not been identified.

Ceramide phosphoethanolamine, which is an integral part of *Drosophila* cell membranes, is likely inactivated in conditions with low manganese [5**]. Additionally, since MnSOD helps reduce the amount of free radicals, low levels of Mn is thought to cause damage to cells due to increased oxidative stress; however, excessively high levels of Mn are known to also increase oxidative stress [31]. These examples help illustrate that Mn deficiency as well as excess Mn levels are damaging to various cellular activities of insects. However, in twenty-three species belonging to two different fly families (*Tephritidae* and *Drosophilidae*), no differences in Mn accumulation were observed between the families, suggesting that strong homeostatic mechanisms for metal control are present in insects [32]. The nature of such regulation for Mn at the cellular or systemic level remains unknown.

Effects of Mn on behavior

In social insects, environmental exposure to Mn has been associated with alterations in social tasks, as worker honeybees had higher concentrations of manganese in their body as compared to bees with other tasks for the colony [30**]. Importantly, Mn supplementation in the beehive altered the distribution of bees from defenders of the hive to foragers [30**]. Mn concentration within honey is used as an indicator of Mn in the environment [33,34]. Mn-treated flies also had a significant increase in acetylcholinesterase (AChE) activity [35] and a simultaneous decrease in dopamine levels and tyrosine hydroxylase activity [35]. In mammals, Mn was shown to accumulate in a region of the brain where dopamine syntheses is high [36,37]. Synchrotron X-ray fluorescence microscopy techniques in insects could be used to examine the differential location of Mn within the body and within specific tissues [38–40].

Physiological effects of Mn

Few studies have directly investigated the effects of Mn on physiological functions of tissues and systems in insects. Considering that Mn^{2+} can block Ca^{2+} flux in skeletal muscle in cockroaches [41], it is not surprising that muscle excitation and contraction are also affected in skeletal, smooth, and cardiac muscle function in other insects. This is demonstrated by studies indicating that the hindgut of the cockroach stops contracting after exposure to 2 mM Mn^{2+} [42] and that the heartbeat of *Drosophila* larvae drastically reduces when exposed to 2.5 mM and is eliminated at 15 mM $MnCl_2$ or $MnSO_4$ [43**]. The effects of manganese on locomotion and skeletal muscle function are a combination of effects, likely involving Mn^{2+} blocking voltage-gated Ca^{2+} channels in the presynaptic nerve terminals [43**]. Thus, muscles will show reduced contraction. The body wall muscle in larval *Drosophila* have graded the excitatory junction potentials, which is related to graded contractions, and demonstrate a dose-dependent depression with Mn [43**]. Additionally, acute exposure of 2.5 $MnSO_4$ mM blocks nerve evoked synaptic transmission [43**]. Depending on the ion channel subtypes and density of channels in a neuron, a Mn block of Ca^{2+} influx may even result in a depolarized state if a constitutively active calcium-activated potassium channel ($K_{(Ca)}$) is compromised. If this occurs, then a membrane potential may not repolarize as quickly and result in overexcitation of a neuron and synaptic responses if there is still enough Ca^{2+} influx for synaptic transmission to take place [44]. This may explain the hyperexcitability of crustacean sensory neurons when a nerve is bathed in low Ca^{2+} [45]. However, the action potential amplitude of cockroach motor neurons recorded with an intracellular electrode was compromised by 40 mM Mn^{2+} , which slightly enhanced the amplitude and prolonged the width of the action potential [46]. This supports the notion that Mn^{2+} can reduce a $K_{(Ca)}$ current

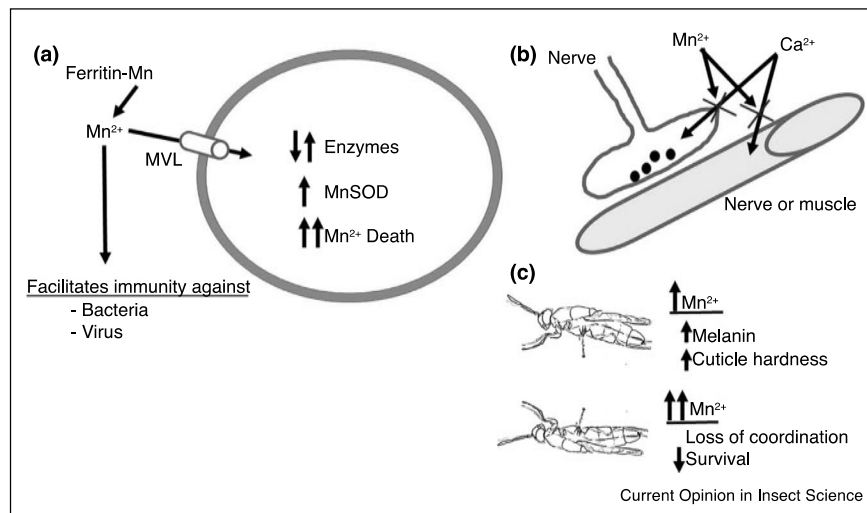
by blocking Ca^{2+} channels. The body wall muscle in most insects has Ca^{2+} channels on the plasmalemma of the muscle to allow Ca^{2+} influx for muscle contraction when depolarized. The direct actions of Mn on muscle, independent of neural innervation, can have consequences due to voltage-gated Ca^{2+} channels on the plasma membrane, essential to electrical depolarization [43**,47]. However, the depolarization of the larval body wall muscle fibers in a beetle (*Xylotrupes dichotomus*) were maintained in the presence of 40 mM Mn^{2+} [48], suggesting the Ca^{2+} channels present were not blocked by Mn, but instead manganese was able to permeate through the channels. The role of Ca^{2+} association in promoting various transport processes are also likely affected by Mn exposure. However, vitellogenin uptake in a cockroach was shown not to be inhibited by Mn [49].

Toxicity effects of Mn

Considering Mn has a role in maintaining an effective immune response, low Mn levels can reduce the life span by causing insects to be more vulnerable to bacterial, viral and parasitic infections. If cellular processes are disturbed in various tissues, it is reasonable to expect that there would also be behavioral consequences from altered Mn levels. Since primary sensory neurons, synaptic transmission at neuromuscular junction and cardiac function are all dampened in response to high Mn levels, it is not surprising that life span may be compromised. Various studies have investigated the mortality rate after Mn exposure in *Drosophila*. The fruit fly is a model organism commonly utilized for pathological conditions related to mammals [50–54]. One study exposed adult *Drosophila* to 10 mM $MnCl_2$ and examined locomotor behaviors as well as survival and showed about 20% mortality after 5 days and nearly 100% mortality after 20 days [55]. The increase in mortality correlated with a rise in reactive oxygen species and a decrease in locomotor behaviors [55]. In another study, adult *Drosophila* exposed to 30 mM/Kg in their diet for seven days resulted in a 20% mortality [56]. The effects of 5 mM $MnCl_2$ or $MnSO_4$ in food eaten by adult *Drosophila* were similar to controls, but 15 mM and 30 mM significantly increased mortality. Eight days of feeding 30 mM $MnSO_4$ resulted in 100% mortality; feeding 30 mM $MnCl_2$ for the same duration was less severe as some animals survived [43**]. $MnSO_4$ or $MnCl_2$ at 30 and 100 mM are toxic to 1st instar *Drosophila* larvae; at 15 mM of either compound, there was still significant mortality (15 mM $MnSO_4$ resulted in 50% of the larvae pupating and eclosing as adults, [43**]). A behavioral analysis of larvae consuming Mn 30 mM $MnSO_4$ or $MnCl_2$ for 24 hours resulted in significant decreases in body wall and mouth hook contractions; 15 mM Mn had a milder but still significant effect [43**].

When comparing freshwater midges, *C. javanus* was equally or more sensitive to environmental metals than

Figure 1



Overview of the effects of manganese on cells, tissue, and the whole animal. **(a)** Labile Mn^{2+} may be transported and potentially regulated in hemolymph by ferritin [64]. The proton-coupled metal ion transporter expressed by the *Mvl* gene has been suggested to transport Mn^{2+} into cells. Increases in Mn^{2+} can be beneficial in helping to maintain an increased immune response to bacteria and viruses. Mn^{2+} concentration within cells can result in some enzymes increasing in expression, while others decrease. In particular, MnSOD increases in expression. However, high levels of intracellular Mn^{2+} can trigger apoptosis and cell death. **(b)** Mn^{2+} can block voltage-gated Ca^{2+} channels on presynaptic nerve terminals as well as on muscles (i.e. body wall and cardiac). **(c)** Dietary Mn^{2+} promotes melanin production to protect from solar radiation and increases cuticle hardness. High levels of dietary Mn^{2+} lead to reduced locomotory function and decreased survival.

Chironomus javanus [57*]. The nymphs of a dragonfly (*Tramea cophysa*) were less sensitive than two tropical ostracod (Crustacea) species (*Chlamydotheca* sp. and *Strandesia trispinosa*) to various metal with Mn being the least toxic of the four metals (i.e. $Cd > Hg > Cu > Mn$) studied [58*].

Mn neurotoxicity

Mn as well as iron (Fe) and copper (Cu) are associated with Parkinson's disease [59**]. Likewise, Mn, along with Zinc (Zn), Cu, aluminum (Al) influence the severity of Friedreich's ataxia [60]. *Drosophila* may serve as a useful model to study the effects of metals on movement disorders, since Mn exposure damages dopaminergic neurons in the central nervous system of *Drosophila melanogaster* [59**]. Alterations in the dopaminergic neural circuits affect locomotion in *Drosophila melanogaster* [59**,61]. It is not yet known if Mn is directly affecting the dopamine receptors or exerting its effects through a specific cellular process. Since larval heart rate is also influenced by dopamine it is possible that Mn might alter heart rate by blocking dopamine receptors [62] as well as other cellular processes. The further exploration of the effects in various insect models needs to continue in order to facilitate understanding the effects of Mn poisoning in other animals [63].

A general overview in the impact of Mn at a cellular level to the whole animal is schematically illustrated in Figure 1.

Conflict of interest statement

Nothing declared.

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References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

- of special interest
- of outstanding interest

1. Howe P, Malcolm H, Dobson S: *Manganese and its Compounds: Environmental Aspects*. World Health Organization; 2004
Extensive review in where manganese occurs in the environment. The ionic forms of manganese in different environments and potential exposures to biological systems.
2. Hakala M, Rantamaki S, Puputti EM, Tyystjarvi T, Tyystjarvi E: **Photoinhibition of manganese enzymes: insights into the mechanism of photosystem II photoinhibition**. *J Exp Bot* 2006, 57:1809-1816.
3. Alejandro S, Höller S, Meier B, Peiter E: **Manganese in plants: from acquisition to subcellular allocation**. *Front Plant Sci* 2020, 11:300.

4. Korcak RF: **Response of blueberry species to excessive manganese.** *J Am Soc Horticult Sci* 1988, **113**:189-193.
5. Vásquez-Procopio J, Osorio B, Cortés-Martínez L, Hernández-Hernández F, Medina-Contreras O, Ríos-Castro E, Comjean A, Li F, Hu Y, Mohr S *et al.*: **Intestinal response to dietary manganese depletion in *Drosophila*.** *Metalomics* 2020, **12**:218-240
 This is a unique study in which a specially defined media was made to deplete *Drosophila* of Mn. Manganese depletion reduced SOD2 activity. In addition, other manganese-dependent enzymatic activities were investigated. A decrease in ceramide phosphoethanolamine synthase activity is key as it is a structural component of cellular membranes.
6. Duttaroy A, Meidinger R, Kirby K, Carmichael S, Hilliker A, Phillips J: **A manganese superoxide dismutase-encoding cDNA from *Drosophila melanogaster*.** *Gene* 1994, **143**:223-225.
7. Flückiger S, Scapozza L, Mayer C, Blaser K, Folkers G, Cramer R: **Immunological and structural analysis of IgE-mediated cross-reactivity between manganese superoxide dismutases.** *Int Arch Allergy Immunol* 2002, **128**:292-303.
8. Xikeranmu Z, Abdunaser M, Ma J, Tusong K, Liu X:
 - **Characterization of two copper/zinc superoxide dismutases (Cu/Zn-SODs) from the desert beetle *Microdera punctipennis* and their activities in protecting *E. coli* cells against cold.** *Cryobiology* 2019, **87**:15-27
 This study demonstrated that cold stress increased the expression of Mn-SOD in a beetle. The tissue of the hindgut was the highest on mRNA levels and was also shown to be present in other regions of the gut and fat bodies as well as the head. The lowest level of expression was found in the head.
9. Tu XH, Zhuo YL, Jiang XY, Liu HL, Cao Y, Liu S: **Molecular**
 - **characterization of a mitochondrial manganese superoxide dismutase from *Chilo suppressalis* (Lepidoptera: Crambidae).** *J Econ Entomol* 2018, **111**:2391-2400
 The authors cover the function of SOD to protect cells against oxidative stress. The authors showed mitochondrial manganese SOD has four manganese binding residues and the expression was present at all developmental stages, but highest in pupae of the Asiatic rice borer. In addition, that the expression level of MnSOD can vary depending on environmental temperatures.
10. Wang Y, Oberley LW, Murhammer DW: **Antioxidant defense**
 - **systems of two lipidopteran insect cell lines.** *Free Radic Biol Med* 2001, **30**:1254-1262
 Examining insect derived cell lines (Tn-5B1-4 and Sf-9) contain MnSOD. The Tn-5B1-4 line MnSOD was about two thirds of the total SOD activity. The Sf-9 cell line had a high level of catalase (CAT) activity. Neither cell line had glutathione peroxidase common in mammalian cells.
11. Wang Y, Oberley LW, Howe D, Jarvis DL, Chauhan G, Murhammer DW: **Effect of expression of manganese superoxide dismutase in baculovirus-infected insect cells.** *Appl Biochem Biotechnol* 2004, **119**:181-193.
12. Hoek JB, Pearson DJ, Olembo NK: **Nicotinamide-adenine dinucleotide-linked "malic" enzyme in flight muscle of the tsetse fly (*Glossina*) and other insects.** *Biochem J* 1976, **160**:253-262.
13. Devorshak C, Roe RM: **Purification and characterization of a phosphoric triester hydrolase from the tufted apple bud moth, *Platynota idaeusalis* (Walker).** *J Biochem Mol Toxicol* 2001, **15**:55-65.
14. Alizamani T, Shakarami J, Mardani-Talaei M, Zibaei A, Serrão JE: **Micronutrient fertilizers affect the digestibility, intermediary metabolism, and oxidative stress in *Myzus persicae* (Sulzer).** *Neotrop Entomol* 2021, **50**:940-947.
15. Newsholme EA, Crabtree B, Higgins SJ, Thornton SD, Start C: **The activities of fructose diphosphatase in flight muscles from the bumble-bee and the role of this enzyme in heat generation.** *Biochem J* 1972, **128**:89-97.
16. Vadaie N, Hulinsky RS, Jarvis DL: **Identification and characterization of a *Drosophila melanogaster* ortholog of human beta1,4-galactosyltransferase VII.** *Glycobiol* 2002, **12**:589-597.
17. Kulkarni GV, Deobagkar DD: **A cytosolic form of aminopeptidase P from *Drosophila melanogaster*: molecular cloning and characterization.** *J Biochem* 2002, **131**:445-452.
18. Fan CL, Brown GM: **Partial purification and properties of guanosine triphosphate cyclohydrolase from *Drosophila melanogaster*.** *Biochem Genet* 1976, **14**:259-270.
19. Mertsalov IB, Novikov BN, Scott H, Dangott L, Panin VM: **Characterization of *Drosophila* CMP-sialic acid synthetase activity reveals unusual enzymatic properties.** *Biochem J* 2016, **473**:1905-1916.
20. Hildenbrandt GR, Bieber LL: **Characterization of glycerophosphorylcholine, -ethanolamine, -serine, -inositol, and -glycerol hydrolytic activity in housefly larvae.** *J Lipid Res* 1972, **13**:348-355 PMID: 4337155.
21. Fritz MA, Hotchkiss PG, Fallon AM: **Changes in ribonuclease activity during development of the mosquito, *Aedes aegypti*.** *Comp Biochem Physiol B* 1986, **84**:355-361.
22. Koyama T, Matsubara M, Ogura K: **Isoprenoid enzyme systems of silkworm. II. Formation of the juvenile hormone skeletons by farnesyl pyrophosphate synthetase II.** *J Biochem* 1985, **98**:457-463.
23. Ogawa H, Nagamura Y, Ishiguro I: **Cinnabaric acid formation in Malpighian tubules of the silkworm, *Bombyx mori*. Participation of catalase in cinnabaric acid formation in the presence of manganese ion.** *Hoppe Seylers Z Physiol Chem* 1983, **364**:1059-1066.
24. Morishima I: **Cyclic nucleotide phosphodiesterase in silkworm. Characterization of cyclic GMP phosphodiesterase.** *Biochim Biophys Acta* 1975, **410**:310-317.
25. Morishima I: **Adenylate cyclase in silkworm. Properties of the enzyme in pupal fat body.** *J Biochem* 1978, **84**:1495-1500.
26. Jeng WY, Wang NC, Lin MH, Lin CT, Liaw YC, Chang WJ, Liu CI, Liang PH, Wang AH: **Structural and functional analysis of three β -glucosidases from bacterium *Clostridium cellulovorans*, fungus *Trichoderma reesei* and termite *Neotermes koshunensis*.** *J Struct Biol* 2011, **173**:46-56.
27. Alexander SPH, Kelly E, Mathie A, Peters JA, Veale EL, Armstrong JF, Faccenda E, Harding SD, Pawson AJ, Sharman JL *et al.*: **CGTP collaborators: the concise guide to pharmacology 2019/20: transporters.** *Br J Pharmacol* 2019, **176**:S397-S493.
28. Orgad S, Nelson H, Segal D, Nelson N: **Metal ions suppress the**
 - **abnormal taste behavior of the *Drosophila* mutant malvolio.** *J Exp Biol* 1998, **201**:115-120
 A mutation in the mvl gene decreasing the transport of Mn affects taste behavior in *Drosophila melanogaster*. Feeding high levels of MnCl₂ recovered the normal taste choice. Interestingly, adult mutant flies to mvl only took 2 hours of feeding MnCl₂ was sufficient to restore normal taste behavior.
29. Šovik E, LaMora A, Seehra G, Barron AB, Duncan JG, Ben-Shahar Y: ***Drosophila* divalent metal ion transporter Malvolio is required in dopaminergic neurons for feeding decisions.** *Genes Brain Behav* 2017, **16**:506-514
 The Mvl gene expresses the only defined natural resistance-associated macrophage protein (NRAMP) in *Drosophila*. The Mvl related protein transports Mn into cells. The authors have demonstrated that Mvl expression in the dopaminergic neurons is important food choice. Dietary Mn helps to maintain food choices.
30. Ben-Shahar Y, Dudek NL, Robinson GE: **Phenotypic**
 - **deconstruction reveals involvement of manganese transporter malvolio in honey bee division of labor.** *J Exp Biol* 2004, **207**:3281-3288
 The authors explain the transport of Mn relates to the expression of the mvl gene. The mRNA level of mvl in the head of bees were higher in pollen foragers compared with nurses. Feeding of Mn increased honeybees to feed on sucrose. This study demonstrated that Mn in the diet altered the responsiveness and division of labor in insects.
31. Mora M, Bonilla E, Medina-Leendertz S, Bravo Y, Arcaya JL: **Minocycline increases the activity of superoxide dismutase and reduces the concentration of nitric oxide, hydrogen peroxide and mitochondrial malondialdehyde in manganese**

- treated *Drosophila melanogaster*. *Neurochem Res* 2014, **39**:1270-1278.
32. Rempoulakis P, Afshar N, Osorio B, Barajas-Aceves M, Szular J, Ahmad S, Dammalage T, Tomas US, Nemny-Lavy E, Salomon M *et al.*: **Conserved metallomics in two insect families evolving separately for a hundred million years.** *Biometals* 2014, **27**:1323-1335.
 33. Ćirić J, Spirić D, Baltić T, Lazić IB, Trbović D, Parunović N, Petronijević R, Đorđević V: **Honey bees and their products as indicators of environmental element deposition.** *Biol Trace Elem Res* 2021, **199**:2312-2319.
 34. Silva AS, Araújo SB, Souza DC, Silva FA: **Study of the Cu, Mn, Pb and Zn dynamics in soil, plants and bee pollen from the region of Teresina (PI), Brazil.** *An Acad Bras Cienc* 2012, **84**:881-889.
 35. Silva NC, Poetini MR, Bianchini MC, Almeida FP, Dahle MMM, Araujo SM, Bortolotto VC, Musachio EAS, Ramborger BP, Novo DR *et al.*: **Protective effect of gamma-oryzanol against manganese-induced toxicity in *Drosophila melanogaster*.** *Environ Sci Pollut Res Int* 2021, **28**:17519-17531.
 36. Baek SY, Lee MJ, Jung HS, Kim HJ, Lee CR, Yoo C, Lee JH, Lee H, Yoon CS, Kim YH *et al.*: **Effect of manganese exposure on MPTP neurotoxicities.** *NeuroToxicology* 2003, **24**:657-665.
 37. Bowman AB, Kwakye GF, Hernandez EH, Aschner M: **Role of manganese in neurodegenerative diseases.** *J Trace Elem Med Biol* 2011, **25**:191-203.
 38. Knebel D, Assaf Y, Ayali A: **The use of MEMRI for monitoring central nervous system activity during intact insect walking.** *J Insect Physiol* 2018, **108**:48-53.
 39. Keim CN, Cruz-Landim C, Carneiro FG, Farina M: **Ferritin in iron containing granules from the fat body of the honeybees *Apis mellifera* and *Scaptotrigona postica*.** *Micron* 2002, **33**:53-59.
 40. Jones MW, de Jonge MD, James SA, Burke R: **Elemental mapping of the entire intact *Drosophila* gastrointestinal tract.** *J Biol Inorg Chem* 2015, **20**:979-987.
 41. Wegener C, Nässel DR: **Peptide-induced Ca(2+) movements in a tonic insect muscle: effects of proctolin and periviscerokinin-2.** *J Neurophysiol* 2000, **84**:3056-3066.
 42. Cook BJ, Holman GM: **The role of proctolin and glutamate in the excitation-contraction coupling of insect visceral muscle.** *Comp Biochem Physiol C Comp Pharmacol Toxicol* 1985, **80**:65-73.
 43. Pankau C, Nadolski J, Tanner H, Cryer C, Di Girolamo J, Haddad C, Lanning M, Miller M, Neely D, Wilson R *et al.*: **Examining the effect of manganese on physiological processes: Invertebrate models.** *Comp Biochem Physiol C Toxicol Pharmacol* 2022, **251**:109209
- This study investigated the effects of MnCl₂ and MnSO₄ on survival, development, cardiac function, and motor function of larval *Drosophila*. The effects of Mn²⁺ on synaptic transmission at the neuromuscular junction were also addressed in larval *Drosophila* and crayfish models. The study also analyzed effects on sensory nerves, where Mn²⁺ was shown to depress activity of proprioceptive nerves in a marine crab.
44. Faber ES, Sah P: **Calcium-activated potassium channels: multiple contributions to neuronal function.** *Neuroscientist* 2003, **9**:181-194.
 45. Atkins DE, Bosh KL, Breakfield GW, Daniels SE, Devore MJ, Fite HE, Guo LZ, Henry DKJ, Kaffenberger AK, Manning KS *et al.*: **The effect of calcium ions on mechanosensation and neuronal activity in proprioceptive neurons.** *NeuroSci* 2021, **2**:353-371.
 46. Pitman RM: **The ionic dependence of action potentials induced by colchicine in an insect motoneuron cell body.** *J Physiol* 1975, **247**:511-520.
 47. Washio H: **The ionic requirements for the initiation of action potentials in insect muscle fibers.** *J Gen Physiol* 1972, **59**:121-134.
 48. Fukuda J, Kawa K: **Permeation of manganese, cadmium, zinc, and beryllium through calcium channels of an insect muscle membrane.** *Science* 1977, **196**:309-311.
 49. Kindle H, Lanzrein B, Kunkel JG: **The effect of ions, ion channel blockers, and ionophores on uptake of vitellogenin into cockroach follicles.** *Dev Biol* 1990, **142**:386-391.
 50. Kim T, Song B, Lee IS: ***Drosophila* glia: models for human neurodevelopmental and neurodegenerative disorders.** *Int J Mol Sci* 2020, **21**:4859.
 51. Higgins J, Hermanns C, Malloy C, Cooper RL: **Considerations in repetitive activation of light sensitive ion channels for long term studies: channel rhodopsin in the *Drosophila* model.** *Neurosci Res* 2017, **125**:1-10.
 52. Akasaka T, Ocorr K: **Drug discovery through functional screening in the *Drosophila* heart.** *Methods Mol Biol* 2009, **577**:235-249.
 53. Bellen HJ, Tong C, Tsuda H: **100 years of *Drosophila* research and its impact on vertebrate neuroscience: a history lesson for the future.** *Nat Rev Neurosci* 2010, **11**:514-522.
 54. Xiao G: **Determination of metal content in *Drosophila melanogaster* during metal exposure.** *Methods Mol Biol* 2021, **2326**:327-337.
 55. Oboh G, Ogunsuyi OB, Awonyemi OI, Atoki VA: **Effect of alkaloid extract from African Jointfir (*Gnetum africanum*) leaves on manganese-induced toxicity in *Drosophila melanogaster*.** *Oxid Med Cell Longev* 2018, **2018**:8952646.
 56. Adedara IA, Abolaji AO, Rocha JB, Farombi EO: **Diphenyl diselenide protects against mortality, locomotor deficits and oxidative stress in *Drosophila melanogaster* model of manganese-induced neurotoxicity.** *Neurochem Res* 2016, **41**:1430-1438.
 57. Shuhaimi-Othman M, Yakub N, Umirah NS, Abas A: **Toxicity of eight metals to Malaysian freshwater midge larvae *Chironomus javanus* (Diptera, Chironomidae).** *Toxicol Ind Health* 2011, **27**:879-886
- Larvae of freshwater midge *Chironomus javanus* were exposed over four days at controlled levels. Mortality was assessed and median lethal concentrations (LC(50)) were calculated. LC(50)s for 96 hours was conducted with a ranking was Cd & Cu & Fe & Pb & Al & Mn & Zn & Ni in toxicity. Comparison of LC(50) values for metals for this species with those for other freshwater midges reveals that *C. javanus* is equally or more sensitive to metals than most other tested dipteran.
58. César Dos Santo sLima J, Gazonato Neto AJ, de Pádua Andrade D, Freitas EC, Moreira RA, Miguel M, Daam MA, Rocha O: **Acute toxicity of four metals to three tropical aquatic invertebrates: the dragonfly *Tramea cophysa* and the ostracods *Chlamydotheca sp.* and *Strandesia trispinosa*.** *Ecotoxicol Environ Saf* 2019, **180**:535-541
- The aim of this study was to evaluate the acute toxicity of four metals (Cd, Cu, Mn, and Hg) in a tropical dragonfly nymph of *Tramea cophysa* and two tropical ostracod species (*Chlamydotheca sp.* and *Strandesia trispinosa*). The order of metal sensitivity was different for the three test species: *T. cophysa*: Cu & CdHg & Mn, *Chlamydotheca sp.*: Cd & Cu & Hg & Mn, and *S. trispinosa*: Cd & Hg & Cu & Mn. However, manganese was the least toxic metal tested for all three species, which is hypothesized to be due to a possible metal transfer to the cuticle of the moulting test species.
59. Bonilla-Ramirez L, Jimenez-Del-Rio M, Velez-Pardo C: **Acute and chronic metal exposure impairs locomotion activity in *Drosophila melanogaster*: a model to study Parkinsonism.** *Biometals* 2011, **24**:1045-1057
- These authors showed that acute (15 mM for up to five days) or chronic (0.5 mM for up to 15 days) exposure to Fe, Mn and Cu reduced life span and locomotor activity in adult fruit flies (*Drosophila melanogaster*). The metals accumulated in the fly's head and were capable to destroy dopamine-ergic neurons. This work provides for the first time metal-induced Parkinson-like symptoms in *D. melanogaster*.
60. Soriano S, Calap-Quintana P, Llorens JV, Al-Ramahi I, Gutiérrez L, Martínez-Sebastián MJ, Botas J, Moltó MD: **Metal homeostasis regulators suppress frda phenotypes in a *Drosophila* model of the disease.** *PLoS One* 2016, **11**:e0159209.
 61. Yamamoto S, Seto ES: **Dopamine dynamics and signaling in *Drosophila*: an overview of genes, drugs and behavioral paradigms.** *Exp Anim* 2014, **63**:107-119.

62. Titlow JS, Rufer JM, King KE, Cooper RL: **Pharmacological analysis of dopamine modulation in the *Drosophila melanogaster* larval heart.** *Physiol Rep* 2013, 1:e00020.
63. Bianchini MC, Gualarte CO, Escoto DF, Pereira G, Gayer MC, Roehrs R, Soares FA, Puntel RL: ***Peumus boldus* (Boldo) aqueous extract present better protective effect than boldine against manganese-induced toxicity in *D. melanogaster*.** *Neurochem Res* 2016, 41:2699-2707.
64. Gutiérrez L, Zubow K, Nield J, Gambis A, Mollereau B, Lázaro FJ, Missirlis F: **Biophysical and genetic analysis of iron partitioning and ferritin function in *Drosophila melanogaster*.** *Metallomics* 2013, 5:997-1005.